www.rsc.org/obc

Oxidation of *N*-substituted dopamine derivatives: irreversible formation of a spirocyclic product

Edward J. Land,^a Almudena Perona,^a Christopher A. Ramsden^{*a} and Patrick A. Riley^b

 ^a School of Chemistry and Physics, Keele University, Keele, Staffordshire, UK ST5 5BG. E-mail: c.a.ramsden@chem.keele.ac.uk; Fax: 01782 712378; Tel: 01782 583045
^b Grav Cancer Institute, Mount Vernon Hospital, Northwood, UK HA6 2JR

Received 29th April 2005, Accepted 10th May 2005

First published as an Advance Article on the web 19th May 2005

Oxidation of amide, urea and guanidinium derivatives of dopamine gives relatively stable *ortho*-quinones whereas oxidation of corresponding thioamide and amidinium derivatives rapidly and quantitatively gives novel bicyclic and spirocyclic products formed *via* the corresponding *ortho*-quinone.

Using a combination of tyrosinase oximetry, pulse radiolysis and chemical oxidation we have studied the transformations of a variety of novel *ortho*-quinone amines **2**, formed by chemical or enzymatic oxidation of the corresponding catechols.¹ These studies have shown that the final product can vary considerably depending upon the structure of the catechol precursor and the lifetimes of intermediate species (Scheme 1).²⁻⁶ Oxidation of higher homologues of dopamine gives transient spirocyclic products **1** (n = 1, 2) but these kinetic products rapidly re-open leading to the thermodynamically more stable *para*-quinomethanes **3** or the benzoheterocycles **4**. In this communication we report the irreversible formation of a novel and stable spirocyclic product by oxidation of an amidine derivative of dopamine.



In vertebrates, tyrosinase oxidation of dopa leads to formation of the melanin pigments.⁷⁻⁹ In insects, oxidation of *N*-acetyldopamine **5** (NADA) leads to enzyme-assisted isomerisation of the *ortho*-quinone **7** to the corresponding *para*quinomethane, which is a reactive intermediate involved in the sclerotization of insect cuticle.¹⁰ In the search for a dopamine derivative related to NADA **5** that may either give the isomeric quinomethane (*cf.* **3**) without enzyme assistance or act as a prototype tyrosinase-activated prodrug¹¹ by cyclising to form a readily hydrolysable intermediate (*cf.* **4**), we have studied a series of closely related derivatives, including compounds **6** and **9–11**. These derivatives were selected to give a wide variation of



 pK_a and nucleophilicity of the side-chain functional group. The catechols **5**¹² and **10**¹³ were prepared by literature methods. The catechols **6**, **9** and **11** were prepared by demethylation (aq. 48% HBr at 90 °C) of the corresponding dimethyl ethers^{14–16} and were purified and fully characterised before oxidation.

Oxidation of the catechols **5** (amide $pK_a \approx -1$) and **6** (urea $pK_a \approx +1$) in CDCl₃/CH₃OH solution using DDQ (1 equiv.) resulted in rapid formation of the *ortho*-quinones **7** and **8** (3 aromatic H, δ 6.2–7.2). These products are relatively stable and slowly decompose over two days to give complex mixtures, including some unreacted quinone. Similar behaviour was observed in aqueous solution using pulse radiolytic or tyrosinase oxidation.

In contrast, oxidation (DDQ) of the thiourea **9** (thiourea $pK_a \approx -1$) resulted in rapid quantitative formation of a product that was not the *ortho*-quinone **12** since only two protons were observed in the aromatic region of the ¹H NMR spectrum. This product was isolated as the hydrochloride salt (mp 227–228 °C), after recrystallisation from ethanolic HCl, and was identified as the 8,9-dihydro-5-thia-7-aza-benzocycloheptene **13** [δ 3.07 (2H, t, *J* 6.0 Hz), 3.13 (3H, s), 3.69 (2H, t, *J* 6.0 Hz), 6.59 (1H, s), 6.66 (1H, s), 31.3 (CH₂), 32.4 (NCH₃), 46.8 (NCH₂), 115.1 (C), 118.6 (CH), 119.8 (CH), 133.4 (C), 146.2 (C), 149.3 (C), 167.3 (C)]. This heterocycle has a UV spectrum (λ_{max} 287 nm) identical



to the product formed by tyrosinase oxidation of catechol 9, and pulse radiolysis showed that product 13 is formed ($k = 1.8 \text{ s}^{-1}$ at pH 8) from the *ortho*-quinone 12 (λ_{max} 380 nm).

When the amidine **10** (amidine $pK_a \approx 11-12$) was oxidised (1 equiv. DDQ) an NMR spectrum was obtained that appeared to be consistent with quantitative formation of the *ortho*-quinone. However, this product was remarkably more stable

than the ortho-quinones 7 and 8 and the ¹H NMR spectrum was unchanged after several days. This product was isolated as colourless crystals, mp 144 °C (decomp.) [M⁺ (cation) m/z294.9802 (${}^{35}Cl$) = $C_{10}H_{10}N_2O_2Cl_3$]. Closer inspection of the ¹H and ¹³C NMR spectra revealed this product to have the spirocyclic structure 15. In particular, the ¹H NMR spectrum shows two signals due to the methylene protons (δ 2.20 and 3.85). The signal at δ 3.85 (t) is entirely consistent with the NCH₂ function. The signal at δ 2.20 is too high field to be a benzylic signal (δ 2.75 in **10**) but is entirely consistent with a spirocyclic CH₂ attached to a quaternary carbon. Furthermore, this signal appears as a quartet due to the non-equivalence of the methylene protons. The structural assignment 15 is further supported by the ¹³C NMR spectrum which shows signals at δ 29.1 and 38.5, due to the methylene carbons, and a signal a δ 55.3, due to the spiro carbon atom. All other aspects of the NMR spectra support the hexadienone structure 15 [δ 5.95 (1H, d, J 3 Hz), 6.35 (1H, d, J 10 Hz), 7.00 (1H, dd, J 3 and 10 Hz), 87.9 (CCl₃), 115.1, 128.8, 147.5, 149.4 (4C, diene C), 159.0 (N=C-N), and 182.0 (C=O)]. The UV spectrum [λ_{max} (0.1 M phosphate buffer: pH 7.4) 220, 244 (ε 7415) and 320sh (ε 1440) nm] is entirely consistent with the dienone structure and comparable to those observed for the transient species 1 (λ_{max} 250 nm).^{3,5}

In contrast to the amidine **10**, DDQ oxidation of the guanidine **11** (p $K_a \approx 13$) gave a ¹H NMR spectrum [δ 6.50 (1H, s), 6.57 (1H, d, J 7 Hz), 7.30 (1H, d, J 7 Hz)] very similar to those of the amide **5** and the urea **6**, and which is clearly due to the *ortho*-quinone **16**. There was no evidence of cyclisation to the spiro derivative **18**. This observation suggests that it is the equilibrating amidine free base **14** that cyclises ($k \approx 2.5 \times 10^2 \text{ s}^{-1}$ at pH 8) and that the guanidinium cation is too weak an acid to be able to cyclise *via* the free base. If the initial amidine cyclisation product is the 1-proto cation **17**, this will rapidly equilibrate to the resonance stablised 3-proto cation **15** (Scheme 2). Since the transformation **17** to **15** is irreversible this may account for the stability of the spirocyclic species **15**. In the case of the amine spirocycles **1** any





free amine formed can readily reprotonate the nitrogen atom, enabling facile reversal of the cyclisation.

When the NMR solution of the bromide 16 was monitored after four hours it was found to contain an approximately equimolar mixture of the *ortho*-quinone 16 and its catechol precursor 11. The regeneration of the catechol 11 is consistent with the bromide ion reducing $(2Br^- \rightarrow Br_2^{*-} \rightarrow Br_2)$ the *ortho*-quinone (Q) to the semi-quinone (Q^{*-}) followed by rapid disproportionation $(2Q^{*-} \rightarrow Q + Q^{2-})$.

In conclusion, relatively simple but unexplored derivatives of dopamine quinone show an interesting variety of reaction modes. The *ortho*-quinone **14** provides an unusual example of formation of a stable spiro-cyclic product (**15**), which are often kinetic^{3,5} but not thermodynamic products of quinone amine cyclisations. None of the derivatives studied showed evidence of rapid *para*-quinomethane formation assisted by the neighbouring functional group.⁴ The lack of cyclisation of the urea **8** suggests that ureas, carbamates and related species are unlikely to function as prodrugs activated by tyrosinase to form hydrolysable species of the type **4** (R¹ = H, R² = CO.R).¹¹

Acknowledgements

We thank the EPSRC National Mass Spectrometry Service Centre for mass spectra, the Ministerio de Educacion y Ciencia, Spain for an FPI Grant (to A. P.), Merck (Harlow, UK) for a contribution to the cost of materials and John Clews (Keele) for technical assistance.

Notes and references

- 1 E. J. Land, C. A. Ramsden and P. A. Riley, Acc. Chem. Res., 2003, 36, 300.
- 2 C. J. Cooksey, P. J. Garratt, E. J. Land, C. A. Ramsden and P. A. Riley, *Biochem. J.*, 1998, **333**, 685.
- 3 J. Clews, C. J. Cooksey, P. J. Garratt, E. J. Land, C. A. Ramsden and P. A. Riley, J. Chem. Soc., Perkin Trans. I, 2000, 4306.
- 4 E. J. Land, C. A Ramsden, P. A. Riley and G. Yoganathan, Org. Biomol. Chem., 2003, 1, 3120.
- 5 E. J. Land, C. A. Ramsden, P. A. Riley and G. Yoganathan, *Pigm. Cell Res.*, 2003, **16**, 397.
- 6 E. J. Land, C. A. Ramsden, P. A. Riley and G. Yoganathan, *Tetrahedron*, 2003, **59**, 9547.
- 7 G. Prota, *Melanins and Melanogenesis*, Academic Press, San Diego, CA, 1992.
- 8 E. J. Land, C. A. Ramsden and P. A. Riley, in *The Pigmentary System: Physiology and Pathophysiology*, J. J. Nordlund, R. E. Boissy, V. J. Hearing, R. A. King, W. S. Oetting and J.-P. Ortonne, eds., Blackwell Publishers, Oxford, 2nd edn., in press.
- 9 E. J. Land, C. A. Ramsden and P. A. Riley, *Methods Enzymol.*, 2004, **378A**, 88.
- 10 M. Sugumaran, Pigm. Cell Res., 2002, 15, 2.
- 11 A. M. Jordan, T. H. Khan, H. M. I. Osborn, A. Photiou and P. A. Riley, *Bioorg. Med. Chem.*, 1999, **7**, 1775.
- 12 Y. Niederstein and M. G. Peter, Liebigs Ann. Chem., 1989, 1189.
- 13 W. S. Saari, M. B. Freedman, J. R. Huff, S. W. King, A. W. Raab, S. J. Bergstrand and E. L. Engelhardt, J. Med. Chem., 1978, 21, 1283.
- 14 J. S. Buck, J. Am. Chem. Soc., 1934, 56, 1607.
- 15 C. G. Stuckwisch and J. S. Hilliard, J. Med. Chem., 1965, 8, 734.
- 16 J. H. Short, U. Biermacher, D. A. Dunnigan and T. D. Leth, J. Med. Chem., 1963, 6, 275.